

Virtual Friday Meetings (EST) Weekly Meetings (Tuesday/Thursday) EST

6/9/21 -- 8:00-8:50 P.M.	8/19/21 -- 8:00-8:40 A.M.
6/11/21 -- 9:00-10:00 P.M.	8/24/21 -- 8:00-8:40 A.M.
6/18/21 -- 9:00-10:00 P.M.	8/26/21 -- 8:00-8:40 A.M.
6/25/21 -- 9:00-10:00 P.M.	8/31/21 -- 8:00-8:40 A.M.
7/2/21 -- 9:00-9:30 P.M.	9/2/21 -- 8:00-8:40 A.M.
7/9/21 -- 9:00-10:00 P.M.	9/7/21 -- 8:00-8:40 A.M.
7/16/21 -- 9:00-10:00 P.M.	9/9/21 -- 8:00-8:40 A.M.
7/23/21 -- 9:00-10:00 P.M.	9/14/21 -- 8:00-8:40 A.M.
7/30/21 -- 9:00-10:00 P.M.	9/16/21 -- 8:00-8:40 A.M.
8/6/21 -- 9:00-10:00 P.M.	9/21/21 -- 8:00-8:40 A.M.
8/20/21 -- 9:00-9:30 P.M.	9/23/21 -- 8:00-8:40 A.M.
8/27/21 -- 9:00-9:45 P.M.	9/28/21 -- 8:00-8:40 A.M.
9/3/21 -- 9:00-9:30 P.M.	9/30/21 -- 8:00-8:40 A.M.
9/10/21 -- 9:00-10:00 P.M.	10/5/21 -- 8:00-8:40 A.M.
9/17/21 -- 9:00-10:00 P.M.	10/7/21 -- 8:00-8:40 A.M.
9/24/21 -- 9:00-10:00 P.M.	10/12/21 -- 8:00-8:40 A.M.
10/1/21 -- 9:00-10:00 P.M.	10/14/21 -- 8:00-8:40 A.M.
10/8/21 -- 9:00-10:00 P.M.	10/19/21 -- 8:00-8:40 A.M.
10/15/21 -- 9:00-10:00 P.M.	10/21/21 -- 8:00-8:40 A.M.

Summer Wet-Lab Meetings (EST)**Tasks Completed**

6/10/21 -- 12:30-2:45 P.M. (Neha, Ahmad)	Prepare E. coli cultures, work on grant
6/11/21 -- 12:20-3:00 P.M. (Neha, Ahmad, Kate)	Making new agar for plates
6/17/21 -- 11:00 A.M.-3:00 P.M. (Neha, Ahmad)	Making MCF tubes with different E. coli cells and phases, MCC procedure, making tubes with different variants (sand, soil, water saturated or not, etc.)
6/18/21 -- 12:20-3:00 P.M. (Neha, Ahmad, Kate)	Transformations, plating
6/21/21 -- 12:30-3:30 P.M. (Neha, Brooke)	Picking cultures, making competent cells, transformations
6/22/21 -- 9:50 A.M.-1:50 P.M. (Neha, Brooke, Kate)	Plasmid prep, plating for tap water + E. coli dilutions, control for C strand, gel + electrophoresis
6/24/21 -- 11:10 A.M. -2:10 P.M. (Neha, Brooke, Kate, Ahmad)	Dilutions, plating, growing plaques for phi-x on E. coli C., ideas for logo, research for project ideas
6/28/21 -- 10:00-2:30 P.M. (Neha, Brooke)	Updating the BioBrick registry for parts, reading papers, making agar, new K-12 and W cultures, new cells and phage, new Amp and K plates (poured all)
6/29/21 -- 12:20-2:30 & 3:30-4:00 P.M. (Neha, Brooke)	Making competent cells, new DNA, doing transformations and plating
7/1/21 -- 10:00-12:00 & 1:00-3:00 P.M. (Neha, Brooke)	Picking colonies, restocking and sterilizing tips and micro tubes, making

	lysates, K-12 and W cell cultures, and plating.
7/2/21 -- 10:00-1:00 P.M. (Neha, Brooke)	Pouring plates, counting colonies, plasmid preps and running plasmid, digests, planning/brainstorming promotion video ideas
7/5/21 -- 10:00-3:00 P.M. (Neha, Kate)	Plasmid prep, running plasmid digests, making and loading gel, freezing down E. coli cells, making agar with antibiotics/pouring plates, making competent cells, making control cells and plating (on plain plates)
7/7 /21 -- 10:00-12:00 & 1:30-3:00 P.M. (Neha, Brooke, Kate)	Doing transformations (pulled DNA), liquid culture, making broth, plating, planning for video
7/8/21 -- 10:00-1:00 P.M. (Neha, Brooke, Kate)	Picking colonies, plating to test phages with E. coli strains, planning video
7/9/21 -- 10:00-3:00 P.M. (Neha, Brooke, Kate)	Plasmid preps, digest for sets 1 and 2, running plasmid
7/12/21 -- 12:20-3:20 P.M. (Neha, Brooke, Kate)	Making video (filming), editing and putting together video clips
7/19/21 -- 10:20-3:20 P.M. (Neha, Ahmad, Brooke, Kate)	Testing phages, writing procedures into Benchling, making agar and soft agar
7/21/21 -- 11:20-12:00 P.M. (Neha, Brooke)	Pouring plates, restocking lab materials, analyzing plates and phages, making plan and procedure for future wet lab work

7/22/21 -- 12:20-2:20 P.M. (Neha, Ahmad, Brooke, Kate)	E. Coli dilutions, working on video submission
7/23/21 -- 10:30-12:30 P.M. (Neha, Brooke)	Submitted video, uploading procedures and data to online notebook, sending emails and papers, pouring "chlor" plates
7/26/21 -- 11:30-11:40 P.M. (Neha, Brooke, Kate)	Plating E. coli and phages, pulling and analyzing plates
7/27/21 -- 11:00-11:50 & 1:40-3:40 P.M. (Neha, Brooke, Kate)	Picking colonies, making E. Coli and phages, plating
7/28/21 -- 11:00-2:30 P.M. (Neha, Brooke, Kate)	Making more phages and competent cells, pouring plates, updating Benchling
7/29/21 -- 12:30-1:00 P.M. (Neha, Brooke, Kate)	Phage and cell plates, collected pond water samples
7/30/21 -- 11:30-2:30 P.M. (Neha, Ahmad, Brooke, Kate)	Dilutions, plating phages and cells, risk assessment study
8/5/21 -- 10:00-5:30 P.M. (Kate, Eric)	MidAtlantic meetup
8/11/21 -- 11:00-3:00 P.M. (Neha, Ahmad, Brooke, Kate)	Looking for wild phages around Gaston Day School's Campus
9/7/21 -- 8:00-8:40 P.M. (Neha)	Dilution of phages with W strand
9/9/21 -- 8:00-8:30 P.M. (Neha, Brooke)	Grew new K-12, W, and A culture, working on BioBrick registry parts
10/1/21 -- 10/15/21 (Neha, Brooke, Kate)	Filling in the lab-work gaps, consolidating results data, and working on the wiki

Physical Notebook Entries

06/23/21

C-D C-C white/clear
 W-F D-E

Digest of EcoRS

5x DNB

Mol. Weight
 EcoRS 10000
 DNB 5000

10 x 11 = 110
 5 x 11 = 55

NO VIBRATIONS
 K-12 10ML 9: 12.001
 K-12 10ML 4: 249.001
 K-12 10ML 6: 0.007

2019/2020 Physical Progs

1. C. 10000
 2. C. 10000
 3. C. 10000
 4. C. 10000
 5. C. 10000
 6. C. 10000
 7. C. 10000
 8. C. 10000
 9. C. 10000
 10. C. 10000

2019/2020 Physical Progs

1. C. 10000
 2. C. 10000
 3. C. 10000
 4. C. 10000
 5. C. 10000
 6. C. 10000
 7. C. 10000
 8. C. 10000
 9. C. 10000
 10. C. 10000

Genetic Progs 7/15/21

Genetic Progs 7/15/21

1. C. 10000
 2. C. 10000
 3. C. 10000
 4. C. 10000
 5. C. 10000
 6. C. 10000
 7. C. 10000
 8. C. 10000
 9. C. 10000
 10. C. 10000

Genetic Will Progs: 7/15/21

1. Plasmid prepared from previously grown plates (P1 or P2)
 2. General and strain into LB near final
 3. Standard mabing and phase procedures
 in 100 uL each cell (100, 10, 1)

Notes to Will Progs from step 2
 1. incubate at 37C for 15 mins
 2. mix with melted top agar and pour into plain plates

Will Progs:

Cell	W	1
Cell	W	2
Cell	W	3
Cell	W	4
Cell	W	5
Cell	W	6
Cell	W	7
Cell	W	8
Cell	W	9
Cell	W	10

Growing Phages (Re-check) 7/20/21

1 ml cell
 +
 100 ul wild phage

1. incubate at 37C for 15 mins
 2. mix with melted top agar and pour into Chlor. plates

Wild Phages:

Cell	W	1
Cell	W	2
Cell	W	3
Cell	W	4
Cell	W	5
Cell	W	6
Cell	W	7
Cell	W	8
Cell	W	9
Cell	W	10

Genetic Will Progs

1. C. 10000
 2. C. 10000
 3. C. 10000
 4. C. 10000
 5. C. 10000
 6. C. 10000
 7. C. 10000
 8. C. 10000
 9. C. 10000
 10. C. 10000

K12 + W dilutions

1 2 3 4 5 6
 100 100 100 100 100 100

K12:

100:100 = 100%
 100:100 = 100%
 100:100 = 100%

W:

100:100 = 100%
 100:100 = 100%
 100:100 = 100%

Redo Dilution

1 2 3 4 5 6
 100 100 100 100 100 100

W:

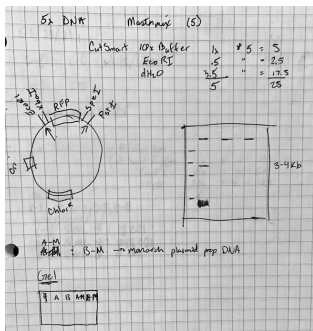
100:100 = 100%
 100:100 = 100%
 100:100 = 100%

Null From BioBrick Registry

1. E114062 GFP Terminator 2008 UK AMP RFP 6
 2. E114061 GFP Terminator 2008 UK AMP RFP 6
 3. E114060 GFP Terminator 2008 UK AMP RFP 6
 4. E114059 GFP Terminator 2008 UK AMP RFP 6
 5. E114058 GFP Terminator 2008 UK AMP RFP 6
 6. E114057 GFP Terminator 2008 UK AMP RFP 6
 7. E114056 GFP Terminator 2008 UK AMP RFP 6
 8. E114055 GFP Terminator 2008 UK AMP RFP 6
 9. E114054 GFP Terminator 2008 UK AMP RFP 6
 10. E114053 GFP Terminator 2008 UK AMP RFP 6

jit. resistance sort

A:0 K12 1 RFP - Chlor 1
 A:1 K12 2 RFP - Chlor 2
 A:2 K12 3 RFP - Chlor 3
 A:3 K12 4 RFP - Chlor 4
 A:4 K12 5 RFP - Chlor 5
 A:5 K12 6 RFP - Chlor 6
 A:6 K12 7 RFP - Chlor 7
 A:7 K12 8 RFP - Chlor 8
 A:8 K12 9 RFP - Chlor 9
 A:9 K12 10 RFP - Chlor 10



Null From BioBrick Registry

1. E114062 GFP Terminator 2008 UK AMP RFP 6
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 7. E114056 GFP Terminator 2008 UK AMP RFP 6
 8. E114055 GFP Terminator 2008 UK AMP RFP 6
 9. E114054 GFP Terminator 2008 UK AMP RFP 6
 10. E114053 GFP Terminator 2008 UK AMP RFP 6

jit. resistance sort

A:0 K12 1 RFP - Chlor 1
 A:1 K12 2 RFP - Chlor 2
 A:2 K12 3 RFP - Chlor 3
 A:3 K12 4 RFP - Chlor 4
 A:4 K12 5 RFP - Chlor 5
 A:5 K12 6 RFP - Chlor 6
 A:6 K12 7 RFP - Chlor 7
 A:7 K12 8 RFP - Chlor 8
 A:8 K12 9 RFP - Chlor 9
 A:9 K12 10 RFP - Chlor 10