

# Gel extraction with Quiagen QIAquick

## Notes:

- All centrifugation steps are carried out at 13 000 rpm.

## For sequencing (more thorough washing)

1. Excise the DNA fragment from the agarose gel with a clean, sharp scalpel.
  - How to cut DNA out of gel - using UV imaging in the gel room in floor 3 - remember lab coat, gloves and UV shield!
    1. Make sure you have images your gel
    2. Weigh your tubes without gel
    3. Clean cast and scalpel with ethanol
    4. Put gel on cast
    5. Put on UV shields + turn the blinds + turn off the lights
    6. Cut the gel with 4 incisions as close as possible to the DNA
    7. Image after cutting to see that you have everything
    8. Weigh your tubes with gel
2. Add 3 volumes Buffer QG to 1 volume gel (1 mg gel  $\approx$  1  $\mu$ L). The maximum amount of gel per spin column is 400 mg. For > 2% agarose gels, add 6 volumes Buffer QG
3. Incubate at 50 °C for 10 min. Vortex the tube every 2-3 min to help dissolve the gel.
  - After the gel slice has dissolved completely, check that the colour of the mixture is yellow (similar to buffer QG without dissolved agarose. If the colour of the mixture is orange or violet, add 10  $\mu$ L 3 M sodium acetate, pH 5.0, and mix. The mixture turns yellow.
4. Add 1 gel volume isopropanol to the sample and mix
5. Place a QIAquick spin column in a provided 2 mL collection tube. To bind DNA, apply the sample to the QIAquick column and centrifuge for 1 min. Discard flow-through and place the QIAquick column back into the same tube. For sample volumes > 800  $\mu$ L, load and spin again.
6. Add 500  $\mu$ L Buffer QG to the QIAquick column and centrifuge for 1 min. Discard flow-through and place the QIAquick column back into the same tube.
7. To wash, add 750  $\mu$ L Buffer PE to the QIAquick column and let the column stand for 2-5 min. Then centrifuge for 1 min. Discard flow-through and place the QIAquick column back into the same tube.
8. Place the QIAquick column into a clean 1.5 mL microcentrifuge tube.
9. Add 30  $\mu$ L Buffer EB to the center of the QIAquick membrane, let the column stand for 4 min, and then centrifuge for 1 min.